

REVIEW ARTICLE

Roles of 3-nitrotyrosine- and 4-hydroxynonenal-modified brain proteins in the progression and pathogenesis of Alzheimer's disease

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Abstract

Proteins play an important role in normal structure and function of the cells. Oxidative modification of proteins may greatly alter the structure and may subsequently lead to loss of normal physiological cell functions and may lead to abnormal function of cell and eventually to cell death. These modifications may be reversible or irreversible. Reversible protein modifications, such as phosphorylation, can be overcome by specific enzymes that cause a protein to 'revert' back to its original protein structure, while irreversible protein modifications cannot. Several important irreversible protein modifications include protein nitration and HNE modification, both which have been extensively investigated in research on the progression of Alzheimer's disease (AD). From the earliest stage of AD throughout the advancement of the disorder there is evidence of increased protein nitration and HNE modification. These protein modifications lead to decreased enzymatic activity, which correlates directly to protein efficacy and provides support for several common themes in AD pathology, namely altered energy metabolism, mitochondrial dysfunction and reduced cholinergic neurotransmission. The current review summarized some of the findings on protein oxidation related to different stages of Alzheimer's disease (AD) that will be helpful in understanding the role of protein oxidation in the progression and pathogenesis of AD.

Keywords: Alzheimer's disease, mild cognitive impairment, early Alzheimer's disease, protein nitration, lipid peroxidation, 4-hydroxy 2-trans nonenal, proteomics

Introduction

Proteins play an important role in normal structure and function of the cells. Oxidative modification of proteins may lead to loss of normal physiological cell functions and may eventually lead to cell death. The current review summarizes some of the findings on protein oxidation related with different stages of Alzheimer's disease (AD) that may relate to the progression and pathogenesis of AD.

AD is an age-related neurodegenerative disease that is pathologically characterized by the presence of extracellular amyloid plaques, intracellular neurofibrillary tangles (NFT) and loss of synaptic connections. The exact mechanism of AD pathogenesis is not clearly understood; however, mutation of *presenilin-1* (PS-1), *presenilin-2* (PS-2) and *amyloid precursor protein* (APP)

genes has been found to be associated with inherited AD [1,2]. In addition to the above-mentioned genes, other risk factor genes such as allele 4 of the *apolipoprotein E* (APOE), *endothelial nitric oxide synthase-3* and *alpha-2-macroglobulin* have been associated with AD [3,4]. A number of hypotheses have been proposed to explain AD pathogenesis such as the amyloid cascade, excitotoxicity, oxidative stress and inflammation hypothesis and all these are based on the role of amyloid beta-peptide (A β) [5–7].

Considerable evidence suggests a role of oxidative stress in the pathophysiology of AD [6,8,9]. For example, oxidative stress in the AD brain is evidenced by decreased levels of antioxidant enzymes and by increased levels of oxidative stress markers such as protein oxidation (indexed by protein carbonyls and 3-nitrotyrosine

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(3-NT)), lipid peroxidation, DNA oxidation, advanced glycation end-products and reactive nitrogen species (ROS) formation. Further, the role of oxidative stress in AD pathogenesis is supported by studies that showed diminished $A\beta$ (1-42)-induced toxicity in the presence of vitamin E in cell culture [10].

Elevated levels of oxidative stress markers have also been found in the mild cognitive impairment brain (MCI), a condition characterized by loss of recent memory without dementia or significant impairment of other cognitive functions and with no loss of activities of daily living [11]. Many MCI subjects show some of the neuropathological features of AD at autopsy and MCI is considered a transition stage between normal cognition and AD. In addition to histopathological similarities to AD, MCI patients also show genetic similarities such as mutations in allele 4 of *apolipoprotein E*, *presenilin 1* and *amyloid precursor protein* [12,13]. Moreover, increased levels of oxidative stress and nitrosative stress have also been reported in early AD (EAD). EAD is considered as an intermediate condition between MCI and late-stage AD; however, information involving this disease stage is limited due to lack of autopsy material. A detailed study involving AD, MCI and EAD would add to the understanding of AD pathogenesis and progression and may also lead to identification of important biomarkers.

Nitrosative stress in the brain of subjects with AD, MCI and EAD

Free radicals such as nitric oxide, produced during the nitric oxide synthase (NOS)-catalysed conversion of L-arginine to L-citrulline, play an important role in physiological conditions, such as vasodilation, but also can be harmful [14]. In 1998, Ferid Murad, Robert F. Furchgott and Louis Ignarro were awarded the Nobel Prize in physiology or Medicine for the discovery of the signalling properties of NO. However, an increase level of NO is harmful to the brain [15]. In addition to NO, other reactive nitrogen species (RNS), such as peroxynitrite, nitrogen dioxide, etc., also are produced in the body [16,17]. Protein modification via RNS usually results from either the reaction of peroxynitrite ($ONOO^-$), a highly reactive molecule formed from the reaction of nitric oxide with superoxide anion ($O_2^{\cdot-}$), resulting in protein-resident 3-NT (Figure 1) or from the reaction of NO (or N_2O_3) with thiols to form S-nitrosothiols (RSNO) [18–20]. Hence, nitration of the proteins could affect the secondary, tertiary and consequently the quaternary structure (if applicable) and thereby alter function of proteins including those involved in cell signalling, catalysis, cytoskeletal organization and inflammatory response. Such altered functions could have detrimental effects on cell viability [21–27]. Protein nitration is a reversible process and has been suggested to serve as a cellular signal [28,29].

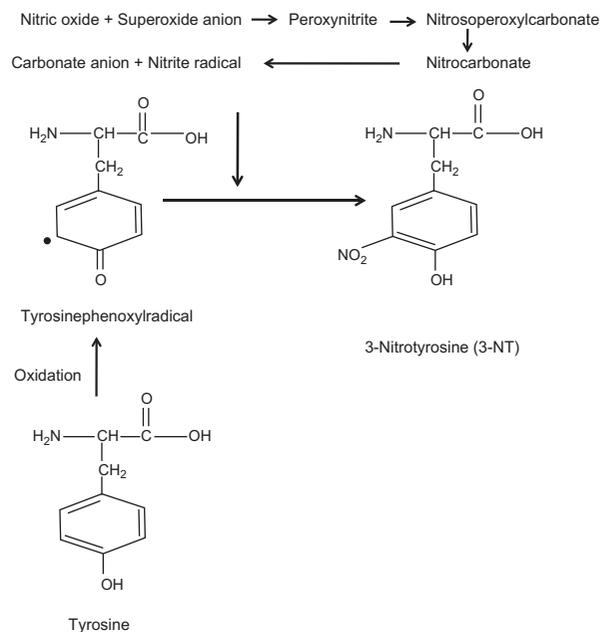


Figure 1. Formation of 3-nitrotyrosine.

Increased levels of RNS lead to increased nitrosative stress that has been reported to play important roles in the pathogenesis of a number of diseases including neurodegenerative disease, ischemia, etc. [30–34]. In AD brain an increase in protein and lipid nitration has been reported [35]. Williamson et al. [36] reported a 2–3-fold increased level of the lipid nitration product, 5-nitro-gamma-tocopherol in AD brain, which correlated with increased NOS levels and thereby suggested a role of nitration in AD pathogenesis [37]. AD brain and ventricular cerebrospinal fluid (VF) showed increased levels of dityrosine (DiTyr) and 3-NT [38] and immunohistochemical analysis showed the presence of nitrated tau in pre-tangles, tangles and tau inclusions in the AD brain. Further, the finding of the increased levels of 3-NT in pre-tangles of early AD brain suggests the involvement of tau nitration as an early event in AD pathogenesis [39,40]. The notion of the protein nitration role in AD was further supported by the fact that the MCI and EAD brain also showed increased levels of protein bound 3-NT [41]. The finding of increased nitration in MCI and EAD brain implies the role of protein nitration in the progression and pathogenesis of AD.

Redox proteomics [42] studies identified a number of proteins to be nitrated in MCI, EAD and AD brains [41]. These proteins were classified based upon their function in various classes (see Table I) and are discussed below with regards to AD progression and pathology.

Energy dysfunction

Of the many proteins that were nitrated in MCI, EAD and AD, only *a*-enolase is found to be a common target of nitration between these different disease stages. This result suggests that *a*-enolase nitration may be

Table I. Functional categorization of nitrated proteins identified in MCI, EAD and AD.

Protein function	MCI ^a	EAD ^d	AD ^{b,c}
Energy dysfunction	<i>α</i>-enolase aldolase malate dehydrogenase	<i>α</i>-enolase phosphoglycerate mutase 1 fructose 1,6- bisphosphate aldolase dehydrogenase triose phosphate isomerase	<i>α</i>-enolase <i>γ</i> -enolase lactate dehydrogenase ^b glyceraldehydes-3- phosphate dehydrogenase triose phosphate isomerase ATP synthase Voltage dependent anion channel protein 1
Mitochondrial dysfunction	—	—	—
Antioxidant defence/detoxification system dysfunction	GSTM3 MRP3 peroxiredoxin VI glucose regulated protein precursor HSPA8 14-3-3- <i>γ</i>	peroxiredoxin II	—
Cell signalling	—	—	—
Lipid abnormalities and cholinergic dysfunction	—	neuropolypeptide h3	—
Neuritic abnormalities and structural dysfunction	DRP2 Fascin 1	—	DRP2 <i>β</i> -actin
Excitotoxicity	—	Glutamate dehydrogenase	—

^aFor nitrated brain proteins identified in MCI see reference [41].

^{b,c}For these nitrated brain proteins identified in AD see references [25,27].

^dFor these nitrated brain proteins identified in EAD see reference [58].

Nitrated proteins **in bold** are common targets of 3-NT modification.

an important event in the progression of AD. Alpha enolase catalyses the conversion of 2-phosphoenolpyruvate to phosphoenolpyruvate in the glycolytic pathway of glucose metabolism and is a frequently oxidatively modified protein in several neurodegenerative diseases including: amyotrophic lateral sclerosis [43], a mouse model of Huntington's disease [44] and a Parkinson's disease mouse model [45,46]. Increased oxidation of alpha-enolase occurs even in models of AD [25,47–51]. In addition to its role in energy metabolism alpha-enolase also perform other functions [52]. Among these are involvements in the regulation of proteolytic clearance of A β and induction of pro-survival pathways. Our laboratory and others have shown that oxidative modification of *α*-enolase in MCI, EAD and AD results in a loss of enzyme function and, hence, in reduced amounts of ATP [53–57]. Elevated chances for neuronal death derived from decreased clearance of A β and diminished induction of ERK1/2-mediated survival pathways also are possible. Consequently, nitration of this protein in MCI, EAD and AD brain would promote progression of AD.

Triose phosphate isomerase (TPI) is a common target of protein nitration between the EAD and AD brain. TPI is a glycolytic enzyme that catalyses isomerization of glyceraldehyde-3-phosphate and dihydroxy acetone phosphate (DHAP). This triose isomerization reaction is important for two reasons: first, it provides

the substrate for the next step of glycolytic cycle and is crucial for ATP production; and, second, by reducing the levels of DHAP TPI prevents the formation of methyl glyoxal, a highly reactive and toxic product. It is interesting to note that the activity of TPI is not altered in the AD or EAD brain [50,58]. TPI deficiencies are associated with hemolytic anaemia, resulting in mitochondrial myopathy [59–61].

Positron emission tomography (PET) studies showed that MCI, EAD and AD brains exhibit decreased glucose utilization compared to age-matched controls. Decreased glucose utilization could be due to impaired functions of the proteins that are involved in the glucose metabolism pathway, which is detrimental to normal brain activity. The stores of ATP at nerve terminals are key for proper neural communication. The decrease in ATP as a consequence of nitration may contribute to synapse loss and dysfunction and, consequently, to the memory impairment and cognitive decline [62] observed in amnesic MCI, EAD and AD. In addition to synapse loss, reduced ATP levels would impair ion-motive ATPase activity with subsequent altered cell potential, loss of membrane lipid asymmetry, intra- and inter-cellular communication and elevated intracellular Ca²⁺ [63,64].

Not all the proteins belonging to energy metabolism pathways that were found to be excessively nitrated in MCI are nitrated in late-stage AD. It is unclear as to what processes reverse nitration of selected proteins.

A recent study suggests bilirubin may play such a role [65]. Bilirubin is a final product of the action of heme oxygenase-1 (HO-1) and the levels of HO-1 and its activity are altered in AD and A β [34,66]. A better understanding of proteins identified that reduce protein nitration may help in development of eventual treatments to slow progression of AD.

Neuritic abnormalities

Axonal growth and repair is crucial for proper neuronal transmission, a key process for memory formation and retrieval. One of the proteins that is important for these functions is dihydropyrimidine-related protein 2 (DRP2), which has been found to be nitrated in both AD and MCI brains [27,41]. DRP2 is involved in neurite outgrowth, so nitration of DRP2 would be predicted to lead to shortened dendritic length, which has been reported in the AD brain [67]. This protein also undergoes other oxidative post-translational modifications, such as protein carbonylation and decreased levels of DRP2 in the AD brain and its animal models have been reported [53,68,69]. Further, individuals with Down syndrome, having an extra copy of *APP* gene and consequently increased A β , also showed decreased levels of DRP2 [70]. Hence, decreased expression and elevated nitration of DRP2 in AD and MCI brains may contribute to shortened neurites with consequent diminution in cognition [67].

Antioxidant defense/detoxification system dysfunction

Peroxiredoxins (Prx) exist in six isoforms, i.e. Prx I, -II, -II, -IV, V and VI, that reduce hydrogen peroxide. Of these isoforms Prx II and Prx VI are found to be significantly nitrated in EAD and MCI, respectively [41,58]. Both these Prx use different electron donors, i.e. thioredoxin and glutathione for II and IV, respectively; however, they catalyse similar functions. Peroxiredoxin II is neuronal-specific and functions also to help regulate the opening of the mitochondrial permeability transition pore [71] and is thereby important in inhibiting apoptosis [71] and promoting neuronal cell survival [72,73]. Prx2 over-expression in cell lines prevents oxidant-induced apoptosis [74]. The levels of Prx2 were reported to be increased in Parkinson's disease [75] as well as AD and in Down's syndrome [76,77]. In the AD brain the increased level of Prx VI was found to be associated with SP and NFT [78]. In addition to reduction of hydrogen peroxide, Prx II also reduces peroxynitrite and is, thereby, important for detoxification of RNS [79]. Hence, loss of Prx activity may lead to increased ROS and RNS that has been reported to be elevated in the AD brain. Prx VI is an efficient antioxidant enzyme that catalyses the reduction of peroxynitrite [80] and is also involved in cell differentiation and apoptosis. Further, it has been

reported that changes in Prx VI activity may also influence phospholipase A2 activity, a protein regulated by peptidyl prolyl *cis/trans* isomerase (Pin 1). In AD and MCI brains Pin 1 has been shown to be down-regulated, oxidatively modified and has reduced activity that could lead to abnormal tau hyperphosphorylation and consequently to NFT formation [81–83]. Further, Prx VI has been shown to form a complex with glutathione-S-transferase [84], so nitration of Prx is an indication that oxidative stress conditions exist in AD and MCI brains and that peroxiredoxin is an important antioxidant enzyme in the human brain defenses system.

HNE-modification of brain proteins in AD, MCI and EAD

Lipid oxidation in the brain readily occurs due to the fact that lipids are particularly vulnerable since polyunsaturated fatty acids are abundant in brain and oxygen is present in membrane bilayers at high levels. Lipid peroxidation is a complex process involving the interaction of oxygen-derived free radicals with polyunsaturated fatty acids, resulting in a variety of highly reactive electrophilic aldehydes that are capable of easily attaching covalently to proteins by forming adducts with cysteine, lysine or histidine residues (Figure 2) through Michael addition [85–88]. Although malondialdehyde and 4-hydroxy-2-nonenal (HNE) represent the major products of lipid peroxidation [85], other carbonyl-containing products are formed, among which are acrolein, neuroprostanes and isoprostanes [89–91].

Lipid peroxidation is highly evident in several neurodegenerative diseases including PD [92], Huntington's disease (HD) [93] and AD [94–97]. Specifically in AD, protein-bound HNE and acrolein have been found to be significantly elevated [98–100]. Evidence

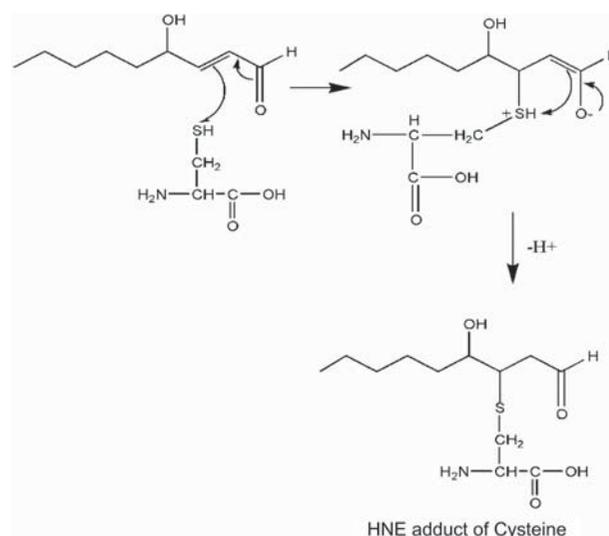


Figure 2. Michael addition of a cysteine residue to HNE.

indicates that lipid peroxidation is an early event during the progression of AD, as demonstrated by its appearance in MCI [101] as well as early AD [102] and late-stage AD [103].

Analysis of AD brains demonstrates an increase in free HNE in amygdala, hippocampus and parahippocampal gyrus of the AD brain compared with age-matched controls [104]. This increased alkenal concentration corresponds with the regions showing the most striking histopathologic alterations in AD. A significant elevation of free HNE in ventricular CSF and serum provides a potential biomarker for AD [105,106]. Likewise, protein-bound HNE, which is an indication of Michael addition of HNE to proteins, is elevated in AD [27,107,108]. Other lipid peroxidation products such as acrolein, malondialdehyde and isoprostanes have been significantly elevated in MCI [102,109,110], EAD [102,110] and late-stage AD [90,91] as well.

Protein-bound HNE alters conformation and function of proteins [107,111–113]. Several important categories of proteins are HNE-modified throughout the course of AD. They include: energy metabolism, mitochondrial dysfunction, cytoskeletal integrity, antioxidant defense, protein synthesis, stress response, neuronal communication and excitotoxicity (Table II).

Energy metabolism

Since altered energy metabolism and reduced cholinergic activity are two well-documented characteristics of AD, it is important to note the HNE modification of several cholinergic, glycolytic and ATP generating

proteins, thereby providing insights into these functions at an early time point of the disease. As noted above, α -enolase catalyses the conversion of 2-phosphoenolpyruvate to phosphoenolpyruvate and is a frequently oxidatively modified protein in several neurodegenerative diseases including: amyotrophic lateral sclerosis [43], a mouse model of Huntington's disease [44] and a Parkinson's disease mouse model [44,45]. α -enolase demonstrates increased oxidation in AD and models of AD [49,53,57,68,69,114]. Protein oxidation and reduced enzyme activity has been previously established in MCI [27], EAD [107] and AD [53,115]. Protein modification of α -enolase may disrupt neuronal energy metabolism and ion homeostasis, thereby impairing the function of membrane ion-motive ATPases and glucose and glutamate transporters [113,116], loss of membrane asymmetry [117] and signal transduction. HNE modification of this protein bolsters the concept that altered energy metabolism is a common theme in neurodegenerative disease. ATP, the energy source of the cell, is extremely important at nerve terminals for normal neural communication. Decreased levels of cellular ATP at nerve terminals may lead to loss of synapses and synaptic function and loss of cell potential that can affect propagation of action potentials, all of which may ultimately contribute to memory loss, potentially correlating evidence of protein modification to memory loss throughout the spectrum of AD. Phosphoglycerate kinase catalyses the conversion of 1,3-bisphosphoglycerate to 3-phosphoglycerate. This reaction undergoes substrate phosphorylation by phosphoryl transfer from 1, 3-bisphosphoglycerate to ADP to produce one molecule

Table II. Functional categorization of HNE-modified proteins identified in MCI, EAD and AD.

Protein function	MCI ^a	EAD ^b	AD ^c
Energy dysfunction	α-enolase phosphoglycerate kinase pyruvate kinase lactate dehydrogenase B	α-enolase aldolase triose phosphate isomerase aconitase	α-enolase
Mitochondrial dysfunction	ATP synthase	ATP synthase manganese superoxide dismutase malate dehydrogenase	ATP synthase manganese superoxide dismutase
Antioxidant defence/ detoxification system dysfunction	carbonyl reductase 1	—	peroxiredoxin VI
Protein synthesis	elongation factor Tu; initiation factor alpha heat shock protein 70 neuropolyptide h3	—	
Stress response			
Lipid abnormalities and cholinergic dysfunction			
Neuritic abnormalities and structural dysfunction		DRP2	DRP2
Excitotoxicity	β-actin		α -tubulin glutamine synthetase

^aFor HNE-modified proteins identified in MCI, see reference [97].

^bFor these HNE-modified proteins identified in EAD, see reference [107].

^cFor these HNE-modified proteins identified in AD, see reference [108].

HNE-modified proteins **in bold** are a common target of modifications.

of ATP. Impairment of this glycolytic enzyme would result in decreased energy production and irreversible downstream effects, such as multi-drug resistance [118]. Pyruvate kinase catalyses the final step in glycolysis, the conversion of phosphoenolpyruvate to pyruvate, with the concomitant transfer of the high-energy phosphate group from phosphoenolpyruvate to ADP, thereby generating ATP. Under aerobic conditions, pyruvate can be transported to the mitochondria, where it enters the TCA cycle and is further metabolized to produce considerably more ATP through oxidative phosphorylation and substrate for brain-requiring glutamate. Additionally, enzymatic activity is reduced, thus suggesting that oxidative modification leads to impairment of protein function [97]. Lactate dehydrogenase B (LDH) reduces pyruvate to lactate by NADH in glycolysis. The generation of NAD⁺ to oxidize glyceraldehyde-3-phosphate in glycolysis is one pertinent reason for the reduction of pyruvate to lactate. Lactate is a substrate for gluconeogenesis and, given that glucose is the major supplier of energy to the brain, proper lactate production is crucial [119]. The enzyme activity of lactate dehydrogenase was significantly reduced in the MCI hippocampus, which further correlates protein dysfunction and enzyme activity impairment. Impairment of this enzyme could initiate reduction of glucose production and creation of excess pyruvate. Aldolase (ALDO1) cleaves fructose 1,6-bisphosphate into dihydroxyacetone phosphate (DHAP) and glyceraldehyde-3-phosphate in glycolysis. This is a critical step as it generates two substrates that will further undergo reactions to produce two molecules of ATP. Consequently, HNE modification results in decreased energy metabolism. Levels of ALDO1 are significantly decreased in the AD hippocampus [120] and PD [121]. Enzyme activity is reduced [122] and impairment can cause increased levels of fructose 1,6- bisphosphate, inhibition of complete glycolysis and ATP depletion. TPI easily isomerizes dihydroxyacetonephosphate (DHAP) to glyceraldehyde-3-phosphate (G3P) in glycolysis. Transformation of DHAP to G3P is imperative to continue glycolysis and for ATP production. ATP is essential in maintaining ATPases, ion motive pumps and potential gradients. In the AD brain, TPI is oxidatively modified as shown by our group in late stage AD [57,123]. These results support the notion that energy metabolism is a critical component in the progression of AD pathogenesis from MCI to EAD and terminal late-stage AD.

Energy metabolism/mitochondrial dysfunction

Aconitase is a TCA cycle enzyme involved in the conversion of citrate to isocitrate via a dehydration–hydration reaction. The TCA cycle generates a higher level of ATP than glycolysis by energetically equivalent molecules

(i.e. GTP, FAD and NAD). Enzymatic activity of aconitase is significantly reduced in the AD brain [108]. Since aconitase is a mitochondrial protein, impairment can lead to mitochondrial dysfunction as observed in several disorders including PD, Friedrich's ataxia and Huntington's disease [124]. Decreased ATP production can lead to voltage-gated channel and ion-motive pump disruption as well as synapse loss, an early event in AD pathology [125]. ATP synthase is a mitochondrial regulating sub-unit of complex V that plays a key role in energy production. This enzyme complex goes through a sequence of coordinated conformational changes of its major sub-units (α , β) to produce ATP. ATP synthase has been previously shown to be HNE-modified in MCI [97], EAD [107] and late-stage AD [108]. Activity for this enzyme is significantly reduced in the aforementioned stages of AD. The oxidation of ATP synthase leads to the inactivation of this mitochondrial complex.

Altered expression and elevated oxidative modification of mitochondrial proteins, functional deficits and lowered activity in different complexes of the ETC are seen in AD [126–130]. These changes, coupled with the changes in complexes I, III and IV, may cause electron leakage from the mitochondria to produce ROS. This action may also affect the mitochondrial proton gradient and its ROS generation by reason of mitochondrial dysfunction suggests a different hypothesis [131,132] for the acknowledged existence of oxidative stress in AD [9,101,133–135].

Additional mitochondrial dysfunction

Superoxide dismutase (SOD) catalyses the conversion of two superoxide anions to hydrogen peroxide and oxygen (Figure 3). Maintenance of this enzyme is critical to achieving oxidative balance; otherwise the cell would be in a continual state of oxidative stress. There are several distinct forms of SOD including Cu/ZnSOD (SOD1), MnSOD (SOD2), NiSOD and FeSOD. MnSOD is located in the mitochondria and thus impairment of this protein can greatly affect the proteasome causing an oxidized protein 'overload' with the inability to correctly ubiquitinate and degrade oxidized proteins. This notion is further bolstered by research stating that oxidative modification of manganese superoxide dismutase inactivates the enzyme [136]. Activity of MnSOD is significantly reduced in EAD compared to the age-matched control brain [107], consistent with this hypothesis.

Malate dehydrogenase (MDH) catalyses the reversible oxidation of malate to oxaloacetate by NAD⁺ in the TCA cycle. MDH links glycolysis to the ETC by transferring NADH to complex I through the



Figure 3. Disproportionation of superoxide catalysed by the enzyme, superoxide dismutase.

malate-aspartate shuttle, thus stimulating ATP production. Activity of MDH increases during ageing [137,138], which can further bolster the hypothesis of mitochondrial dysfunction in AD.

Cytoskeletal integrity

Actin plays a central role in maintaining cellular integrity, morphology and the structure of the plasma membrane. Actin microfilaments stabilize the neuronal membrane cytoskeleton by maintaining the distribution of membrane proteins and segregating axonal and dendritic proteins [139]. In the CNS, actin is distributed widely in neurons, astrocytes and blood vessels [140] and is particularly concentrated in pre-synaptic terminals, dendritic spines and growth cones. Oxidative modification of actin, by HNE, can lead to loss of membrane cytoskeletal structure, decreased membrane fluidity and trafficking of synaptic proteins and mitochondria [141]. Moreover, actin is involved in the elongation of the growth cone and loss of function of actin could play a role in the loss of synapse and neuronal communication documented in AD [142]. α -tubulin is an isoform of tubulin that alternates with β -tubulin to form a prominent cytoskeletal structure, the microtubule. Microtubules, used to transport cargo from the cell centre to the periphery and vice versa are stabilized by tau, which can become hyperphosphorylated and produce neurofibrillary tangles, a characteristic hallmark of AD. Upon HNE conjugation, α -tubulin is structurally modified and microtubules begin to depolymerize [143]. Therefore, cargo, including mitochondria delivery to and from the pre-synaptic terminal, cannot reach their destination comprising the survival of the neuron [144,145].

Antioxidant defence

Carbonyl reductase is an important enzyme that can reduce carbonyl-containing compounds to their resultant alcohols, thereby diminishing protein carbonyl levels (Figure 4). Subsequent malfunction or down-regulation of this enzyme could contribute to the increased protein carbonyls of AD, which because of the polarity

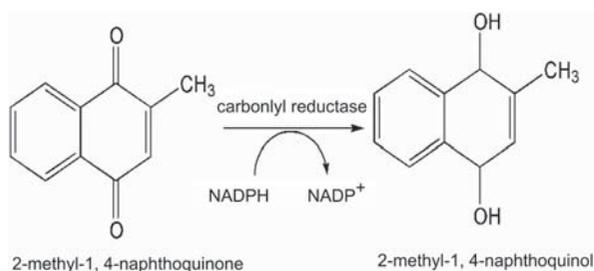


Figure 4. Illustration of one example of a Carbonyl reductase-catalysed reaction.

of the carbonyl moiety could expose ordinarily buried hydrophobic amino acids to the solvent (i.e. disrupt conformation). Carbonyl reductase has been shown to reduce the lipid peroxidation product, HNE [146,147]. Expression of carbonyl reductase is altered in Down's syndrome and AD subjects [148]. The gene for carbonyl reductase is located in close proximity to the gene for Cu/Zn superoxide dismutase (SOD1) [149] and the genes for SOD1, carbonyl reductase and APP are located on chromosome 21, which is a trisomy in Down's syndrome patients [150,151]. A potential association between Down's syndrome and Alzheimer's disease by irregular meiotic recombination in chromosome 21 has been postulated [152]. Our proteomics studies lead us to posit a possible intriguing relationship among $A\beta$, Down syndrome and carbonyl reductase in neurodegeneration [97,107,108].

Peroxiredoxin VI, as stated above, is a nitrated protein and is also found as an HNE-modified protein [108,153]. Prx VI also plays important roles in cell differentiation and apoptosis, so HNE modification may lead to tau hyperphosphorylation and neurofibrillary tangles formation, in addition to development of oxidative stress.

Protein synthesis

Elongation factor Tu and initiation factor a (eIF- a) are intimately involved in protein synthesis machinery. Human mitochondrial EF-Tu (EF-Tu) is a nuclear-encoded protein and functions in the translational apparatus of mitochondria [154]. Mammalian EF-Tu acts as a GTPase and hydrolyses a molecule of GTP each time an amino-acylated tRNA is accommodated on the A site of the ribosome and its recycling depends on the exchange factor EF-Ts. Nuclear genes encode most respiratory chain sub-units and all protein components necessary for maintenance and expression of mtDNA. Mitochondrial protein synthesis inhibition is associated with the impairment of differentiation in different cell types, including neurons [155]. The co-ordination of mitochondrial and nuclear genetic systems in the cell is necessary for proper mitochondrial biogenesis and cellular functioning. eIF- a is an abundant protein required to bind aminoacyl-tRNA to acceptor sites of ribosomes in a GTP-dependent manner during protein synthesis [156]. Eif- a has been shown to be involved in cytoskeletal organization by binding and bundling actin filaments and microtubules. Inhibition of eIF- a induces apoptosis [157], indicating that eIF- a activity is critical to normal cell function. Increased levels of HNE-bound eIF- a and EF-Tu suggest an impairment of the protein synthesis machinery, either in mitochondria or cytosol, associated with an impairment of the rate and specificity of ribosome functions [97]. Numerous studies have provided indirect evidence that suggests alterations in protein synthesis may occur throughout the progression of AD [158–163]. The dysfunction of the protein synthesis apparatus, mediated in part by oxidative stress, could compromise the ability

of cells to generate the various factors needed to regulate cell homeostasis, thus contributing to impaired neuronal function and to the development of neuropathology in AD and pathogenesis of this disorder.

Stress response

Heat shock proteins act as chaperone proteins and aid in protein misfolding, protein aggregation and directing misfolded proteins to the proteasome. Heat shock proteins are involved in combating stress by protecting proteins from denaturation [164]. Heat shock 70 is a member of the heat shock protein family. Several other heat shock proteins have been found to be oxidatively modified in AD [53] and Huntington's disease [44] including Hsp90 and Hsp60 [66]. In the MCI brain, several additional heat shock proteins, including Hsp27, Hsp32 and Hsp70, showed an increase in protein level [66]. Impairment of HSPs may exacerbate protein misfolding and aggregation and eventual proteosomal overload and dysfunction known to occur in AD [165,166]. A β peptide aggregates are the major components of senile plaques, which are a hallmark of AD. A β -treated synaptosomes show that heat shock proteins are oxidatively modified [69], further illustrating the importance of functioning heat shock proteins in the cell.

Neuronal communication

Neuropolypeptide h3 is critical for modulation of the enzyme choline acetyltransferase, which is vital for synthesis of acetylcholine, a neurotransmitter important in signal transduction and cell communication. The loss of choline acetyltransferase leads to reduced levels of acetylcholine causing poor neurotransmission [167]. NMDA receptors activate the production of this enzyme and alteration of the NMDA receptor mediates cholinergic deficits [168]. AD has cholinergic deficits, consistent with dysregulation in acetylcholine levels and loss of cholinergic neurons [169–172]. The HNE modification of neuropolypeptide h3 further supports the involvement of cholinergic neurons, the dysfunction of which is a major neuroclinical deficit in AD [173]. Moreover, cholinergic neurons in the basal forebrain project to the outer molecular layer of the hippocampus, providing a potential link between cholinergic deficits and memory loss in amnesic MCI and AD.

Neuropolypeptide h3 is also known as phosphatidylethanolamine binding protein (PEBP). As a phosphatidylethanolamine binding protein, PEBP could be important in phospholipid asymmetry. A signal for apoptosis to commence is phosphatidylserine becoming exposed to the outer leaflet of the membrane. Loss of function and changes in conformation of PEBP conceivably could lead to loss of phospholipid asymmetry as observed in AD [117,174,175], which can disrupt cellular homeostasis.

As described above, DRP-2 is a member of the dihydropyrimidinase-related protein family that is involved in axonal outgrowth and pathfinding through transmission and modulation of extracellular signals [176,177]. Previous studies reported that DRP-2 can induce growth cone collapse [178,179] by rho-kinase phosphorylation [180] and binding to tubulin heterodimers and bundled microtubule as carriers to promote microtubule assembly and dynamics [181,182]. DRP-2 has been reported to be associated with neurofibrillary tangles, which may lead to decreased levels of cytosolic DRP-2. This, in turn, would eventually lead to abnormal neuritic and axonal growth, thus accelerating neuronal degeneration in AD [183], which is a classic hallmark of AD pathology. Since memory and learning are associated with synaptic remodelling, HNE conjugation and subsequent loss of function of this protein could conceivably be involved in the observed cognitive impairments in this disease. Moreover, the decreased function of DRP-2 could be responsible of shortened dendritic length and synapse loss observed in AD [184]. Shortened dendritic length likely would lead to less neuronal communication with adjacent neurons that could contribute to memory loss and cognitive decline associated with AD.

Excitotoxicity

Glutamine synthetase (GS), an important enzyme in maintaining the glutamate–glutamine cycle, catalyses the conversion of the acidic amino acid, glutamate, to the basic amino acid, glutamine. Glutamate is taken up from extracellular fluid from neuronal tissues via glutamate transporters, particularly EAAT2 (or GLT-1). Once GS is conjugated with HNE, it becomes structurally altered [185] and can no longer preserve glutamate levels, resulting in possible excitotoxicity and neurodegeneration. Glutamine synthetase levels are significantly decreased in the AD brain [186,187], which leads to protein dysfunction and neurodegeneration.

Conclusions

Early diagnosis of AD is essential in delaying the progression of this dementing disease. Hippocampus and IPL are ideal models to use, because they are both damaged in the AD brain. As described above, MCI and EAD are two stages preceding late-stage AD; therefore, especially MCI is a better phase of AD to study in order to gain insights into progression of AD and to potentially identify effective therapeutic agents to slow or halt progression. By using a proteomics-based approach to investigate the oxidative modifications of protein nitration and excessive protein-bound HNE in MCI, EAD and the AD brain, better understanding of the proteins altered during the progression of AD arise. For example, multi-functional α -enolase [52] is a common target protein for both HNE and 3-NT modification, providing a

potential key element in the observed altered energy metabolism, elevated A β and decreased neuronal survival of MCI, EAD and AD. The results of these redox proteomics studies suggest that oxidation of α -enolase might be crucial in the progression of the disease [188–190]. Further, the use of antioxidants or agents that elevate the endogenous levels of cellular defence may help in delaying or preventing this devastating, dementing disorder.

Declaration of Interest

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References

- [1] Shen J, Kelleher RJ, 3rd. The presenilin hypothesis of Alzheimer's disease: evidence for a loss-of-function pathogenic mechanism. *Proc Natl Acad Sci USA* 2007;104:403–409.
- [2] Suh YH, Checler F. Amyloid precursor protein, presenilins, and alpha-synuclein: molecular pathogenesis and pharmacological applications in Alzheimer's disease. *Pharmacol Rev* 2002;54:469–525.
- [3] Levy-Lahad E, Lahad A, Wijsman EM, Bird TD, Schellenberg GD. Apolipoprotein E genotypes and age of onset in early-onset familial Alzheimer's disease. *Ann Neurol* 1995;38:678–680.
- [4] de la Monte SM, Lu BX, Sohn YK, Etienne D, Kraft J, Ganju N, Wands JR. Aberrant expression of nitric oxide synthase III in Alzheimer's disease: relevance to cerebral vasculopathy and neurodegeneration. *Neurobiol Aging* 2000;21:309–319.
- [5] Hardy J, Selkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 2002;297:353–356.
- [6] Butterfield DA, Drake J, Pocernich C, Castegna A. Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid beta-peptide. *Trends Mol Med* 2001;7:548–554.
- [7] Barnham KJ, Ciccosto GD, Tickler AK, Ali FE, Smith DG, Williamson NA, Lam YH, Carrington D, Tew D, Kocak G, Volitakis I, Separovic F, Barrow CJ, Wade JD, Masters CL, Cherny RA, Curtain CC, Bush AI, Cappai R. Neurotoxic, redox-competent Alzheimer's beta-amyloid is released from lipid membrane by methionine oxidation. *J Biol Chem* 2003;278:42959–42965.
- [8] Lauderback CM, Hackett JM, Keller JN, Varadarajan S, Szveda L, Kindy M, Markesbery WR, Butterfield DA. Vulnerability of synaptosomes from apoE knock-out mice to structural and oxidative modifications induced by A beta(1-40): implications for Alzheimer's disease. *Biochemistry* 2001;40:2548–2554.
- [9] Markesbery WR. Oxidative stress hypothesis in Alzheimer's disease. *Free Radic Biol Med* 1997;23:134–147.
- [10] Butterfield DA, Koppal T, Subramaniam R, Yatin S. Vitamin E as an antioxidant/free radical scavenger against amyloid beta-peptide-induced oxidative stress in neocortical synaptosomal membranes and hippocampal neurons in culture: insights into Alzheimer's disease. *Rev Neurosci* 1999;10:141–149.
- [11] Petersen RC, Morris JC. Mild cognitive impairment as a clinical entity and treatment target. *Arch Neurol* 2005;62:1160–1163; discussion 1167.
- [12] Nacmias B, Piccini C, Bagnoli S, Tedde A, Cellini E, Bracco L, Sorbi S. Brain-derived neurotrophic factor, apolipoprotein E genetic variants and cognitive performance in Alzheimer's disease. *Neurosci Lett* 2004;367:379–383.
- [13] Almkvist O, Basun H, Backman L, Herlitz A, Lannfelt L, Small B, Viitanen M, Wahlund LO, Winblad B. Mild cognitive impairment—an early stage of Alzheimer's disease? *J Neural Transm* 1998;(Suppl 54):21–29.
- [14] Lafon-Cazal M, Culcasi M, Gaven F, Pietri S, Bockaert J. Nitric oxide, superoxide and peroxynitrite: putative mediators of NMDA-induced cell death in cerebellar granule cells. *Neuropharmacology* 1993;32:1259–1266.
- [15] Calabrese V, Mancuso C, Calvani M, Rizzarelli E, Butterfield DA, Stella AM. Nitric oxide in the central nervous system: neuroprotection versus neurotoxicity. *Nat Rev Neurosci* 2007;8:766–775.
- [16] Bergendi L, Benes L, Durackova Z, Ferencik M. Chemistry, physiology and pathology of free radicals. *Life Sci* 1999;65:1865–1874.
- [17] Toader V, Xu X, Nicolescu A, Yu L, Bolton JL, Thatcher GR. Nitrosation, nitration, and autoxidation of the selective estrogen receptor modulator raloxifene by nitric oxide, peroxynitrite, and reactive nitrogen/oxygen species. *Chem Res Toxicol* 2003;16:1264–1276.
- [18] Beckman JS, Koppenol WH. Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. *Am J Physiol* 1996;271:1424–1437.
- [19] Broillet MC. S-nitrosylation of proteins. *Cell Mol Life Sci* 1999;55:1036–1042.
- [20] Gow AJ, Buerk DG, Ischiropoulos H. A novel reaction mechanism for the formation of S-nitrosothiol *in vivo*. *J Biol Chem* 1997;272:2841–2845.
- [21] Sampson JB, Rosen H, Beckman JS. Peroxynitrite-dependent tyrosine nitration catalyzed by superoxide dismutase, myeloperoxidase, and horseradish peroxidase. *Methods Enzymol* 1996;269:210–218.
- [22] Koppal T, Drake J, Yatin S, Jordan B, Varadarajan S, Bettenhausen L, Butterfield DA. Peroxynitrite-induced alterations in synaptosomal membrane proteins: insight into oxidative stress in Alzheimer's disease. *J Neurochem* 1999;72:310–317.
- [23] Sennlaub F, Courtois Y, Goureau O. Inducible nitric oxide synthase mediates retinal apoptosis in ischemic proliferative retinopathy. *J Neurosci* 2002;22:3987–3993.
- [24] Ischiropoulos H, Zhu L, Chen J, Tsai M, Martin JC, Smith CD, Beckman JS. Peroxynitrite-mediated tyrosine nitration catalyzed by superoxide dismutase. *Arch Biochem Biophys* 1992;298:431–437.
- [25] Castegna A, Thongboonkerd V, Klein JB, Lynn B, Markesbery WR, Butterfield DA. Proteomic identification of nitrated proteins in Alzheimer's disease brain. *J Neurochem* 2003;85:1394–1401.
- [26] Gow AJ, Duran D, Malcolm S, Ischiropoulos H. Effects of peroxynitrite-induced protein modifications on tyrosine phosphorylation and degradation. *FEBS Lett* 1996;385:63–66.
- [27] Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR, Butterfield DA. Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 2006;22:76–87.
- [28] Koeck T, Fu X, Hazen SL, Crabb JW, Stuehr DJ, Aulak KS. Rapid and selective oxygen-regulated protein tyrosine denitration and nitration in mitochondria. *J Biol Chem* 2004;279:27257–27262.
- [29] Aulak KS, Koeck T, Crabb JW, Stuehr DJ. Dynamics of protein nitration in cells and mitochondria. *Am J Physiol Heart Circ Physiol* 2004;286:30–38.
- [30] Bruijn LI, Beal MF, Becher MW, Schulz JB, Wong PC, Price DL, Cleveland DW. Elevated free nitrotyrosine levels, but not protein-bound nitrotyrosine or hydroxyl radicals, throughout amyotrophic lateral sclerosis (ALS)-like disease implicate tyrosine nitration as an aberrant *in vivo* property of one familial ALS-linked superoxide dismutase 1 mutant. *Proc Natl Acad Sci USA* 1997;94:7606–7611.

- [31] Moncada S, Bolanos JP. Nitric oxide, cell bioenergetics and neurodegeneration. *J Neurochem* 2006;97:1676–1689.
- [32] Kunz A, Park L, Abe T, Gallo EF, Anrather J, Zhou P, Iadecola C. Neurovascular protection by ischemic tolerance: role of nitric oxide and reactive oxygen species. *J Neurosci* 2007;27:7083–7093.
- [33] Malinski T. Nitric oxide and nitroxidative stress in Alzheimer's disease. *J Alzheimers Dis* 2007;11:207–218.
- [34] Calabrese V, Sultana R, Scapagnini G, Guagliano E, Sapienza M, Bella R, Kanski J, Pennisi G, Mancuso C, Stella AM, Butterfield DA. Nitrosative stress, cellular stress response, and thiol homeostasis in patients with Alzheimer's disease. *Antioxid Redox Signal* 2006;8:1975–1986.
- [35] Smith MA, Richey Harris PL, Sayre LM, Beckman JS, Perry G. Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J Neurosci* 1997;17:2653–2657.
- [36] Williamson KS, Gabbita SP, Mou S, West M, Pye QN, Markesbery WR, Cooney RV, Grammas P, Reimann-Philipp U, Floyd RA, Hensley K. The nitration product 5-nitrogamma-tocopherol is increased in the Alzheimer brain. *Nitric Oxide* 2002;6:221–227.
- [37] Fernandez-Vizarrá P, Fernandez AP, Castro-Blanco S, Encinas JM, Serrano J, Bentura ML, Muñoz P, Martínez-Murillo R, Rodrigo J. Expression of nitric oxide system in clinically evaluated cases of Alzheimer's disease. *Neurobiol Dis* 2004;15:287–305.
- [38] Hensley K, Maidt ML, Yu Z, Sang H, Markesbery WR, Floyd RA. Electrochemical analysis of protein nitrotyrosine and dityrosine in the Alzheimer brain indicates region-specific accumulation. *J Neurosci* 1998;18:8126–8132.
- [39] Horiguchi T, Uryu K, Giasson BI, Ischiropoulos H, Lightfoot R, Bellmann C, Richter-Landsberg C, Lee VM, Trojanowski JQ. Nitration of tau protein is linked to neurodegeneration in tauopathies. *Am J Pathol* 2003;163:1021–1031.
- [40] Zhang YJ, Xu YF, Liu YH, Yin J, Li HL, Wang Q, Wang JZ. Peroxynitrite induces Alzheimer-like tau modifications and accumulation in rat brain and its underlying mechanisms. *FASEB J* 2006;20:1431–1442.
- [41] Sultana R, Reed T, Perluigi M, Coccia R, Pierce WM, Butterfield DA. Proteomic identification of nitrated brain proteins in amnesic mild cognitive impairment: a regional study. *J Cell Mol Med* 2007;11:839–851.
- [42] Dalle-Donne I, Scaloni A, Butterfield DA. *Redox proteomics*. New York: Wiley; 2006.
- [43] Ekegren T, Hanrieder J, Aquilonius SM, Bergquist J. Focused proteomics in post-mortem human spinal cord. *J Proteome Res* 2006;5:2364–2371.
- [44] Perluigi M, Poon HF, Maragos W, Pierce WM, Klein JB, Calabrese V, Cini C, De Marco C, Butterfield DA. Proteomic analysis of protein expression and oxidative modification in r6/2 transgenic mice: a model of Huntington disease. *Mol Cell Proteomics* 2005;4:1849–1861.
- [45] De Iuliis A, Grigoletto J, Recchia A, Giusti P, Arslan P. A proteomic approach in the study of an animal model of Parkinson's disease. *Clin Chim Acta* 2005;357:202–209.
- [46] Poon HF, Frasier M, Shreve N, Calabrese V, Wolozin B, Butterfield DA. Mitochondrial associated metabolic proteins are selectively oxidized in A30P alpha-synuclein transgenic mice—a model of familial Parkinson's disease. *Neurobiol Dis* 2005;18:492–498.
- [47] Boyd-Kimball D, Sultana R, Poon HF, Lynn BC, Casamenti F, Pepeu G, Klein JB, Butterfield DA. Proteomic identification of proteins specifically oxidized by intracerebral injection of amyloid beta-peptide (1-42) into rat brain: implications for Alzheimer's disease. *Neuroscience* 2005;132:313–324.
- [48] Castegna A, Aksenov M, Aksenova M, Thongboonkerd V, Klein JB, Pierce WM, Booze R, Markesbery WR, Butterfield DA. Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part I: creatine kinase BB, glutamine synthase, and ubiquitin carboxy-terminal hydrolase L-1. *Free Radic Biol Med* 2002;33:562–571.
- [49] Poon HF, Farr SA, Thongboonkerd V, Lynn BC, Banks WA, Morley JE, Klein JB, Butterfield DA. Proteomic analysis of specific brain proteins in aged SAMP8 mice treated with alpha-lipoic acid: implications for aging and age-related neurodegenerative disorders. *Neurochem Int* 2005;46:159–168.
- [50] Sultana R, Boyd-Kimball D, Poon HF, Cai J, Pierce WM, Klein JB, Merchant M, Markesbery WR, Butterfield DA. Redox proteomics identification of oxidized proteins in Alzheimer's disease hippocampus and cerebellum: an approach to understand pathological and biochemical alterations in AD. *Neurobiol Aging* 2006;27:1564–1576.
- [51] Sultana R, Poon HF, Cai J, Pierce WM, Merchant M, Klein JB, Markesbery WR, Butterfield DA. Identification of nitrated proteins in Alzheimer's disease brain using a redox proteomics approach. *Neurobiol Dis* 2006;22:76–87.
- [52] Butterfield DA, Lange ML. Multifunctional roles of enolase in Alzheimer's disease brain: beyond altered glucose metabolism. *J Neurochem* 2009;111:915–933.
- [53] Castegna A, Aksenov M, Thongboonkerd V, Klein JB, Pierce WM, Booze R, Markesbery WR, Butterfield DA. Proteomic identification of oxidatively modified proteins in Alzheimer's disease brain. Part II: dihydropyrimidinase related protein II, α -enolase and heat shock cognate 71. *J Neurochem* 2002;82:1524–1532.
- [54] Aksenova M, Butterfield DA, Zhang SX, Underwood M, Geddes JW. Increased protein oxidation and decreased creatine kinase BB expression and activity after spinal cord contusion injury. *J Neurotrauma* 2002;19:491–502.
- [55] Meier-Ruge W, Iwagoff P, Reichlmeier K. Neurochemical enzyme changes in Alzheimer's and Pick's disease. *Arch Gerontol Geriatr* 1984;3:161–165.
- [56] Butterfield DA, Poon HF, St. Clair D, Keller JN, Pierce WM, Klein JB, Markesbery WR. Redox proteomics identification of oxidatively modified hippocampal proteins in mild cognitive impairment: insights into the development of Alzheimer's disease. *Neurobiol. Dis* 2006;22:223–232.
- [57] A Castegna VT, Klein JB, Lynn B, Markesbery WR, Butterfield DA. Proteomic identification of nitrated proteins in Alzheimer's disease brain. *J Neurochem* 2003;85:1394–1401.
- [58] Reed TT, Pierce WM, Jr, Turner DM, Markesbery WR, Butterfield DA. Proteomic identification of nitrated brain proteins in early Alzheimer's disease inferior parietal lobule. *J Cell Mol Med* 2009;13:2019–2029.
- [59] Eber SW, Pekrun A, Bardosi A, Gahr M, Krietsch WK, Kruger J, Matthei R, Schroter W. Triosephosphate isomerase deficiency: haemolytic anaemia, myopathy with altered mitochondria and mental retardation due to a new variant with accelerated enzyme catabolism and diminished specific activity. *Eur J Pediatr* 1991;150:761–766.
- [60] Zanella A, Mariani M, Colombo MB, Borgna-Pignatti C, De Stefano P, Morgese G, Sirchia G. Triosephosphate isomerase deficiency: 2 new cases. *Scand J Haematol* 1985;34:417–424.
- [61] Kishi H, Mukai T, Hirono A, Fujii H, Miwa S, Hori K. Human aldolase A deficiency associated with a hemolytic anemia: thermolabile aldolase due to a single base mutation. *Proc Natl Acad Sci USA* 1987;84:8623–8627.
- [62] Hoyer S. Memory function and brain glucose metabolism. *Pharmacopsychiatry* 2003;36:62–67.
- [63] Butterfield DA, Sultana R. Redox proteomics identification of oxidatively modified brain proteins in Alzheimer's disease and mild cognitive impairment: insights into the progression of this dementing disorder. *J Alzheimers Dis* 2007;12:61–72.
- [64] Planel E, Miyasaka T, Launey T, Chui DH, Tanemura K, Sato S, Murayama O, Ishiguro K, Tatebayashi Y, Takashima A. Alterations in glucose metabolism induce hypothermia lead-

- ing to tau hyperphosphorylation through differential inhibition of kinase and phosphatase activities: implications for Alzheimer's disease. *J Neurosci* 2004;24:2401–2411.
- [65] Mancuso C, Barone E. The heme oxygenase/biliverdin reductase pathway in drug research and development. *Curr Drug Metab* 2009;10:579–594.
- [66] Di Domenico F, Sultana R, Tiu GF, Scheff NN, Perluigi M, Cini C, Butterfield DA. Protein levels of heat shock proteins 27, 32, 60, 70, 90 and thioredoxin-1 in amnesic mild cognitive impairment: an investigation on the role of cellular stress response in the progression of Alzheimer disease. *Brain Res* 2010;1333:72–81.
- [67] Coleman PD, Flood DG. Neuron numbers and dendritic extent in normal aging and Alzheimer's disease. *Neurobiol Aging* 1987;8:521–545.
- [68] Sultana R, Boyd-Kimball D, Poon HF, Cai J, Pierce WM, Klein JB, Merchant M, Markesbery WR, Butterfield DA. Redox proteomics identification of oxidized proteins in Alzheimer's disease hippocampus and cerebellum: an approach to understand pathological and biochemical alterations in AD. *Neurobiol Aging* 2006;27:1564–1576.
- [69] Boyd-Kimball D, Castegna A, Sultana R, Poon HF, Petroze R, Lynn BC, Klein JB, Butterfield DA. Proteomic identification of proteins oxidized by Aβ(1–42) in synaptosomes: implications for Alzheimer's disease. *Brain Res* 2005;1044:206–215.
- [70] Lubec G, MN, Krapfenbauer K, Gratzner M, Cairns N, Fountoulakis M. Expression of the dihydropyrimidinase related protein 2 (DRP-2) in Down syndrome and Alzheimer's disease brain is down-regulated at the mRNA and dysregulated at the protein level. *J Neural Transm* 1999;(Suppl 57):161–177.
- [71] Kowaltowski AJ, Vercesi AE, Rhee SG, Netto LE. Catalases and thioredoxin peroxidase protect *Saccharomyces cerevisiae* against Ca(2+)-induced mitochondrial membrane permeabilization and cell death. *FEBS Lett* 2000;473:177–182.
- [72] Zhang P, Liu B, Kang SW, Seo MS, Rhee SG, Obeid LM. Thioredoxin peroxidase is a novel inhibitor of apoptosis with a mechanism distinct from that of Bcl-2. *J Biol Chem* 1997;272:30615–30618.
- [73] Ichimiya S, Davis JG, O'Rourke DM, Katsumata M, Greene MI. Murine thioredoxin peroxidase delays neuronal apoptosis and is expressed in areas of the brain most susceptible to hypoxic and ischemic injury. *DNA Cell Biol* 1997;16:311–321.
- [74] Bae JY, Ahn SJ, Han W, Noh DY. Peroxiredoxin I and II inhibit H(2)O(2)-induced cell death in MCF-7 cell lines. *J Cell Biochem* 2007;101:1038–1045.
- [75] Basso M, Giraud S, Corpillo D, Bergamasco B, Lopiano L, Fasano M. Proteome analysis of human substantia nigra in Parkinson's disease. *Proteomics* 2004;4:3943–3952.
- [76] Kim SH, Fountoulakis M, Cairns N, Lubec G. Protein levels of human peroxiredoxin subtypes in brains of patients with Alzheimer's disease and Down syndrome. *J Neural Transm* 2001;(Suppl):223–235.
- [77] Krapfenbauer K, Engidawork E, Cairns N, Fountoulakis M, Lubec G. Aberrant expression of peroxiredoxin subtypes in neurodegenerative disorders. *Brain Res* 2003;967:152–160.
- [78] Power JH, Asad S, Chataway TK, Chegini F, Manavis J, Temlett JA, Jensen PH, Blumbergs PC, Gai WP. Peroxiredoxin 6 in human brain: molecular forms, cellular distribution and association with Alzheimer's disease pathology. *Acta Neuropathol* 2008;115:611–622.
- [79] Bryk R, Griffin P, Nathan C. Peroxynitrite reductase activity of bacterial peroxiredoxins. *Nature* 2000;407:211–215.
- [80] Peshenko IV, Shichi H. Oxidation of active center cysteine of bovine 1-Cys peroxiredoxin to the cysteine sulfenic acid form by peroxide and peroxynitrite. *Free Radic Biol Med* 2001;31:292–303.
- [81] Lu KP. Phosphorylation-dependent prolyl isomerization: a novel cell cycle regulatory mechanism. *Prog Cell Cycle Res* 2000;4:83–96.
- [82] Lu PJ, Wulf G, Zhou XZ, Davies P, Lu KP. The prolyl isomerase Pin1 restores the function of Alzheimer-associated phosphorylated tau protein. *Nature* 1999;399:784–788.
- [83] Sultana R, Boyd-Kimball D, Poon HF, Cai J, Pierce WM, Klein JB, Markesbery WR, Zhou XZ, Lu KP, Butterfield DA. Oxidative modification and down-regulation of Pin1 in Alzheimer's disease hippocampus: a redox proteomics analysis. *Neurobiol Aging* 2006;27:918–925.
- [84] Ralat LA, Manevich Y, Fisher AB, Colman RF. Direct evidence for the formation of a complex between 1-cysteine peroxiredoxin and glutathione S-transferase pi with activity changes in both enzymes. *Biochem* 2006;45:360–372.
- [85] Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic Biol Med* 1991;11:81–128.
- [86] Fukuda M, Kanou F, Shimada N, Sawabe M, Saito Y, Murayama S, Hashimoto M, Maruyama N, Ishigami A. Elevated levels of 4-hydroxynonenal-histidine Michael adduct in the hippocampi of patients with Alzheimer's disease. *Biomed Res* 2009;30:227–233.
- [87] Sayre LM, Sha W, Xu G, Kaur K, Nadkarni D, Subbanaounder G, Salomon RG. Immunochemical evidence supporting 2-pentylpyrrole formation on proteins exposed to 4-hydroxy-2-nonenal. *Chem Res Toxicol* 1996;9:1194–1201.
- [88] Nadkarni DV, Sayre LM. Structural definition of early lysine and histidine adduction chemistry of 4-hydroxynonenal. *Chem Res Toxicol* 1995;8:284–291.
- [89] Calingasan NY, Uchida K, Gibson GE. Protein-bound acrolein: a novel marker of oxidative stress in Alzheimer's disease. *J Neurochem* 1999;72:751–756.
- [90] Montine KS, Quinn JF, Zhang J, Fessel JP, Roberts LJ, 2nd, Morrow JD, Montine TJ. Isoprostanes and related products of lipid peroxidation in neurodegenerative diseases. *Chem Phys Lipids* 2004;128:117–124.
- [91] Montine TJ, Beal MF, Cudkovic ME, O'Donnell H, Margolin RA, McFarland L, Bachrach AF, Zackert WE, Roberts LJ, Morrow JD. Increased CSF F2-isoprostane concentration in probable AD. *Neurology* 1999;52:562–565.
- [92] Tsang AH, Chung KK. Oxidative and nitrosative stress in Parkinson's disease. *Biochim Biophys Acta* 2009;1792:643–650.
- [93] Perez-De La Cruz V, Elinos-Calderon D, Robledo-Arratia Y, Medina-Campos ON, Pedraza-Chaverri J, Ali SF, Santamaria A. Targeting oxidative/nitrosative stress ameliorates motor impairment, and attenuates synaptic mitochondrial dysfunction and lipid peroxidation in two models of Huntington's disease. *Behav Brain Res* 2009;199:210–217.
- [94] Galasko D, Montine TJ. Biomarkers of oxidative damage and inflammation in Alzheimer's disease. *Biomark Med* 2010;4:27–36.
- [95] Picklo MJ, Montine TJ, Amarnath V, Neely MD. Carbonyl toxicology and Alzheimer's disease. *Toxicol Appl Pharmacol* 2002;184:187–197.
- [96] Neely MD, Montine TJ. CSF lipoproteins and Alzheimer's disease. *J Nutr Health Aging* 2002;6:383–391.
- [97] Reed T, Perluigi M, Sultana R, Pierce WM, Klein JB, Turner DM, Coccia R, Markesbery WR, Butterfield DA. Redox proteomic identification of 4-hydroxy-2-nonenal-modified brain proteins in amnesic mild cognitive impairment: insight into the role of lipid peroxidation in the progression and pathogenesis of Alzheimer's disease. *Neurobiol Dis* 2008;30:107–120.
- [98] Lauderback CM, Hackett JM, Huang FF, Keller JN, Szweda LI, Markesbery WR, Butterfield DA. The glial glutamate transporter, GLT-1, is oxidatively modified by 4-hydroxy-2-nonenal in the Alzheimer's disease brain: the role of Aβ(1–42). *J Neurochem* 2001;78:413–416.

- [99] Lovell MA, Xie C, Markesbery WR. Acrolein is increased in Alzheimer's disease brain and is toxic to primary hippocampal cultures. *Neurobiol Aging* 2001;22:187–194.
- [100] McGrath LT, McGleenon BM, Brennan S, McColl D, Mc IS, Passmore AP. Increased oxidative stress in Alzheimer's disease as assessed with 4-hydroxynonenal but not malondialdehyde. *QJM* 2001;94:485–490.
- [101] Butterfield DA, Lauderback CM. Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid beta-peptide-associated free radical oxidative stress. *Free Radic Biol Med* 2002;32:1050–1060.
- [102] Williams TI, Lynn BC, Markesbery WR, Lovell MA. Increased levels of 4-hydroxynonenal and acrolein, neurotoxic markers of lipid peroxidation, in the brain in mild cognitive impairment and early Alzheimer's disease. *Neurobiol Aging* 2006;27:1094–1099.
- [103] Casado A, Encarnacion Lopez-Fernandez M, Concepcion Casado M, de La Torre R. Lipid peroxidation and antioxidant enzyme activities in vascular and Alzheimer dementias. *Neurochem Res* 2008;33:450–458.
- [104] Markesbery WR, Lovell MA. Four-hydroxynonenal, a product of lipid peroxidation, is increased in the brain in Alzheimer's disease. *Neurobiol Aging* 1998;19:33–36.
- [105] Selley ML, Close DR, Stern SE. The effect of increased concentrations of homocysteine on the concentration of (E)-4-hydroxy-2-nonenal in the plasma and cerebrospinal fluid of patients with Alzheimer's disease. *Neurobiol Aging* 2002;23:383–388.
- [106] Lovell MA, Ehmann WD, Mattson MP, Markesbery WR. Elevated 4-hydroxynonenal in ventricular fluid in Alzheimer's disease. *Neurobiol Aging* 1997;18:457–461.
- [107] Reed TT, Pierce WM, Markesbery WR, Butterfield DA. Proteomic identification of HNE-bound proteins in early Alzheimer disease: insights into the role of lipid peroxidation in the progression of AD. *Brain Res* 2009;1274:66–76.
- [108] Perluigi M, Sultana R, Cenini G, Di Domenico F, Memo M, Pierce WM, Coccia R, Butterfield DA. Redox proteomics identification of 4-hydroxynonenal-modified brain proteins in Alzheimer's disease: role of lipid peroxidation in Alzheimer's disease pathogenesis. *Proteomics Clin Appl* 2009;3:682–693.
- [109] Markesbery WR, Kryscio RJ, Lovell MA, Morrow JD. Lipid peroxidation is an early event in the brain in amnesic mild cognitive impairment. *Ann Neurol* 2005;58:730–735.
- [110] Keller JN, Schmitt FA, Scheff SW, Ding Q, Chen Q, Butterfield DA, Markesbery WR. Evidence of increased oxidative damage in subjects with mild cognitive impairment. *Neurology* 2005;64:1152–1156.
- [111] Hellberg K, Grimsrud PA, Kruse AC, Banaszak LJ, Ohlendorf DH, Bernlohr DA. X-ray crystallographic analysis of adipocyte fatty acid binding protein (aP2) modified with 4-hydroxy-2-nonenal. *Protein Sci* 2010;
- [112] Siems WG, Hapner SJ, van Kuijk FJ. 4-hydroxynonenal inhibits Na(+)-K(+)-ATPase. *Free Radic Biol Med* 1996;20:215–223.
- [113] Subramaniam R, Roediger F, Jordan B, Mattson MP, Keller JN, Waeg G, Butterfield DA. The lipid peroxidation product, 4-hydroxy-2-trans-nonenal, alters the conformation of cortical synaptosomal membrane proteins. *J Neurochem* 1997;69:1161–1169.
- [114] Sultana R, Perluigi M, Butterfield DA. Redox proteomics identification of oxidatively modified proteins in Alzheimer's disease brain and *in vivo* and *in vitro* models of AD centered around Abeta(1–42). *J Chromatogr B Analyt Technol Biomed Life Sci* 2006;833:3–11.
- [115] Castegna A, Lauderback CM, Mohmmad-Abdul H, Butterfield DA. Modulation of phospholipid asymmetry in synaptosomal membranes by the lipid peroxidation products, 4-hydroxynonenal and acrolein: implications for Alzheimer's disease. *Brain Res* 2004;1004:193–197.
- [116] Mattson MP. Modification of ion homeostasis by lipid peroxidation: roles in neuronal degeneration and adaptive plasticity. *Trends Neurosci* 1998;21:53–57.
- [117] Bader Lange ML, Cenini G, Piroddi M, Abdul HM, Sultana R, Galli F, Memo M, Butterfield DA. Loss of phospholipid asymmetry and elevated brain apoptotic protein levels in subjects with amnesic mild cognitive impairment and Alzheimer disease. *Neurobiol Dis* 2008;29:456–464.
- [118] Duan Z, Lamendola DE, Yusuf RZ, Penson RT, Preffer FI, Seiden MV. Overexpression of human phosphoglycerate kinase 1 (PGK1) induces a multidrug resistance phenotype. *Anticancer Res* 2002;22:1933–1941.
- [119] Kida K, Nishio T, Nagai K, Matsuda H, Nakagawa H. Gluconeogenesis in the kidney *in vivo* in fed rats. Circadian change and substrate specificity. *J Biochem* 1982;91:755–760.
- [120] Sultana R, Boyd-Kimball D, Cai J, Pierce WM, Klein JB, Merchant M, Butterfield DA. Proteomics analysis of the Alzheimer's disease hippocampal proteome. *J Alzheimers Dis* 2007;11:153–164.
- [121] Gomez A, Ferrer I. Increased oxidation of certain glycolysis and energy metabolism enzymes in the frontal cortex in Lewy body diseases. *J Neurosci Res* 2009;87:1002–1013.
- [122] Barreiro E, Gea J, Di Falco M, Kriazhev L, James S, Hussain SN. Protein carbonyl formation in the diaphragm. *Am J Respir Cell Mol Biol* 2005;32:9–17.
- [123] Sultana R, Perluigi M, Butterfield DA. Protein oxidation and lipid peroxidation in brain of subjects with Alzheimer's disease: insights into mechanism of neurodegeneration from redox proteomics. *Antioxid Redox Signal* 2006;8:2021–2037.
- [124] Schapira AH. Mitochondrial involvement in Parkinson's disease, Huntington's disease, hereditary spastic paraplegia and Friedrich's ataxia. *Biochim Biophys Acta* 1999;1410:159–170.
- [125] Scheff SW, Price DA, Schmitt FA, Mufson EJ. Hippocampal synaptic loss in early Alzheimer's disease and mild cognitive impairment. *Neurobiol Aging* 2006;27:1372–1384.
- [126] Chen X, Stern D, Yan SD. Mitochondrial dysfunction and Alzheimer's disease. *Curr Alzheimer Res* 2006;3:515–520.
- [127] Hauptmann S, Keil U, Scherping I, Bonert A, Eckert A, Muller WE. Mitochondrial dysfunction in sporadic and genetic Alzheimer's disease. *Exp Gerontol* 2006;41:668–673.
- [128] Reix S, Mechawar N, Susin SA, Quirion R, Krantic S. Expression of cortical and hippocampal apoptosis-inducing factor (AIF) in aging and Alzheimer's disease. *Neurobiol Aging* 2007;28:351–356.
- [129] Kim SH, Vlkolinsky R, Cairns N, Lubec G. Decreased levels of complex III core protein 1 and complex V beta chain in brains from patients with Alzheimer's disease and Down syndrome. *Cell Mol Life Sci* 2000;57:1810–1816.
- [130] Muller WE, Eckert A, Kurz C, Eckert GP, Leuner K. Mitochondrial dysfunction: common final pathway in brain aging and Alzheimer's disease—therapeutic aspects. *Mol Neurobiol* 2010;41:159–171.
- [131] Mancuso M, Coppede F, Migliore L, Siciliano G, Murri L. Mitochondrial dysfunction, oxidative stress and neurodegeneration. *J Alzheimers Dis* 2006;10:59–73.
- [132] Petrozzi L, Ricci G, Giglioli NJ, Siciliano G, Mancuso M. Mitochondria and neurodegeneration. *Biosci Rep* 2007;27:87–104.
- [133] Zhu X, Lee HG, Casadesus G, Avila J, Drew K, Perry G, Smith MA. Oxidative imbalance in Alzheimer's disease. *Mol Neurobiol* 2005;31:205–217.
- [134] Halliwell B. Oxidative stress and neurodegeneration: where are we now? *J Neurochem* 2006;97:1634–1658.
- [135] Butterfield DA, Perluigi M, Sultana R. Oxidative stress in Alzheimer's disease brain: new insights from redox proteomics. *Eur J Pharmacol* 2006;545:39–50.

- [136] Anantharaman M, Tangpong J, Keller JN, Murphy MP, Markesbery WR, Kinningham KK, St Clair DK. Beta-amyloid mediated nitration of manganese superoxide dismutase: implication for oxidative stress in a APPNLH/NLH X PS-1P264L/P264L double knock-in mouse model of Alzheimer's disease. *Am J Pathol* 2006;168:1608-1618.
- [137] Bubber P, Haroutunian V, Fisch G, Blass JP, Gibson GE. Mitochondrial abnormalities in Alzheimer brain: mechanistic implications. *Ann Neurol* 2005;57:695-703.
- [138] Op den Velde W, Stam FC. Some cerebral proteins and enzyme systems in Alzheimer's presenile and senile dementia. *J Am Geriatr Soc* 1976;24:12-16.
- [139] Battaini F, Pascale A, Lucchi L, Pasinetti GM, Govoni S. Protein kinase C anchoring deficit in postmortem brains of Alzheimer's disease patients. *Exp Neurol* 1999;159:559-564.
- [140] Goldman JE. Immunocytochemical studies of actin localization in the central nervous system. *J Neurosci* 1983;3:1952-1962.
- [141] Dalle-Donne I, Carini M, Vistoli G, Gamberoni L, Giustarini D, Colombo R, Maffei Facino R, Rossi R, Milzani A, Aldini G. Actin Cys374 as a nucleophilic target of alpha,beta-unsaturated aldehydes. *Free Radic Biol Med* 2007;42:583-598.
- [142] Masliah E, Mallory M, Hansen L, DeTeresa R, Alford M, Terry R. Synaptic and neuritic alterations during the progression of Alzheimer's disease. *Neurosci Lett* 1994;174:67-72.
- [143] Gadoni E, Olivero A, Miglietta A, Bocca C, Gabriel L. Cytoskeletal modifications induced by 4-hydroxynonenal. *Cytotechnology* 1993;11(Suppl 1):62-64.
- [144] Neely MD, Boutte A, Milatovic D, Montine TJ. Mechanisms of 4-hydroxynonenal-induced neuronal microtubule dysfunction. *Brain Res* 2005;1037:90-98.
- [145] Neely MD, Sidell KR, Graham DG, Montine TJ. The lipid peroxidation product 4-hydroxynonenal inhibits neurite outgrowth, disrupts neuronal microtubules, and modifies cellular tubulin. *J Neurochem* 1999;72:2323-2333.
- [146] Doorn JA, Maser E, Blum A., Claffey DJ, Petersen DR. Human carbonyl reductase catalyzes reduction of 4-oxonon-2-enal. *Biochemistry* 2004;43:13106-13114.
- [147] Oppermann U. Carbonyl reductases: the complex relationships of mammalian carbonyl- and quinone-reducing enzymes and their role in physiology. *Annu Rev Pharmacol Toxicol* 2007;47:293-22.
- [148] Balcz B, Kirchner L, Cairns N, Fountoulakis M, Lubec G. Increased brain protein levels of carbonyl reductase and alcohol dehydrogenase in Down syndrome and Alzheimer's disease. *J Neural Transm Suppl* 2001;61:193-201.
- [149] Lemieux N, Malfoy B, Forrest GL. Human carbonyl reductase (CBR) localized to band 21q22.1 by high-resolution fluorescence in situ hybridization displays gene dosage effects in trisomy 21 cells. *Genomics* 1993;15:169-172.
- [150] Forrest GL, Gonzalez B. Carbonyl reductase. *Chem Biol Interact* 2000;129:21-40.
- [151] Korenberg JR, Bradley C, Disteche CM. Down syndrome: molecular mapping of the congenital heart disease and duodenal stenosis. *Am J Hum Genet* 1992;50:294-302.
- [152] Petronis A. Alzheimer's disease and down syndrome: from meiosis to dementia. *Exp Neurol* 1999;158:403-413.
- [153] Yoshida Y, Yoshikawa A, Kinumi T, Ogawa Y, Saito Y, Ohara K, Yamamoto H, Imai Y, Niki E. Hydroxyoctadecadienoic acid and oxidatively modified peroxiredoxins in the blood of Alzheimer's disease patients and their potential as biomarkers. *Neurobiol Aging* 2009;30:174-185.
- [154] Ling M, Merante F, Chen HS, Duff C, Duncan AM, Robinson BH. The human mitochondrial elongation factor tu (EF-Tu) gene: cDNA sequence, genomic localization, genomic structure, and identification of a pseudogene. *Gene* 1997;197:325-336.
- [155] Vayssiere JL, Cordeau-Lossouarn L, Larcher JC, Basseville M, Gros F, Croizat B. Participation of the mitochondrial genome in the differentiation of neuroblastoma cells. *In Vitro Cell Dev Biol* 1992;28A:763-772.
- [156] Pestova TV, Hellen CU. The structure and function of initiation factors in eukaryotic protein synthesis. *Cell Mol Life Sci* 2000;57:651-674.
- [157] Tome ME, Fiser SM, Payne CM, Gerner EW. Excess putrescine accumulation inhibits the formation of modified eukaryotic initiation factor 5A (eIF-5A) and induces apoptosis. *Biochem J* 1997;328:847-854.
- [158] Chang RC, Wong AK, Ng HK, Hugon J. Phosphorylation of eukaryotic initiation factor-2alpha (eIF2alpha) is associated with neuronal degeneration in Alzheimer's disease. *Neuroreport* 2002;13:2429-2432.
- [159] Ding Q, Markesbery WR, Cecarini V, Keller JN. Decreased RNA, and increased RNA oxidation, in Ribosomes from Early Alzheimer's Disease. *Neurochem Res* 2006;31:705-710.
- [160] Ferrer I. Differential expression of phosphorylated translation initiation factor 2 alpha in Alzheimer's disease and Creutzfeldt-Jakob's disease. *Neuropathol Appl Neurobiol* 2002;28:441-451.
- [161] Li X, An WL, Alafuzoff I, Soininen H, Winblad B, Pei JJ. Phosphorylated eukaryotic translation factor 4E is elevated in Alzheimer brain. *Neuroreport* 2004;15:2237-2240.
- [162] Sajdel-Sulkowska EM, Marotta CA. Alzheimer's disease brain: alterations in RNA levels and in a ribonuclease-inhibitor complex. *Science* 1984;225:947-949.
- [163] Ding Q, Markesbery WR, Chen Q, Li F, Keller JN. Ribosome dysfunction is an early event in Alzheimer's disease. *J Neurosci* 2005;25:9171-9175.
- [164] Calabrese V, Scapagnini G, Colombrita C, Ravagna A., Pennisi G, Giuffrida Stella AM, Galli F, Butterfield DA. Redox regulation of heat shock protein expression in aging and neurodegenerative disorders associated with oxidative stress: a nutritional approach. *Amino Acids* 2003;25:437-444.
- [165] Magrane J, Smith RC, Walsh K, Querfurth HW. Heat shock protein 70 participates in the neuroprotective response to intracellularly expressed beta-amyloid in neurons. *J Neurosci* 2004;24:1700-1706.
- [166] Keller JN, Hanni KB, Markesbery WR. Impaired proteasome function in Alzheimer's disease. *J Neurochem* 2000;75:436-439.
- [167] Ojika K, Tsugu Y, Mitake S, Otsuka Y, Katada E. NMDA receptor activation enhances the release of a cholinergic differentiation peptide (HCNP) from hippocampal neurons *in vitro*. *Brain Res Dev Brain Res* 1998;106:173-180.
- [168] Jouvenceau A, Dutar P, Billard JM. Alteration of NMDA receptor-mediated synaptic responses in CA1 area of the aged rat hippocampus: contribution of GABAergic and cholinergic deficits. *Hippocampus* 1998;8:627-637.
- [169] Davies P, Terry RD. Cortical somatostatin-like immunoreactivity in cases of Alzheimer's disease and senile dementia of the Alzheimer type. *Neurobiol Aging* 1981;2:9-14.
- [170] Davis BM, Mohs RC, Greenwald BS, Mathe AA, Johns CA, Horvath TB, Davis KL. Clinical studies of the cholinergic deficit in Alzheimer's disease. I. Neurochemical and neuroendocrine studies. *J Am Geriatr Soc* 1985;33:741-748.
- [171] Perry EK, Perry RH, Smith CJ, Purohit D, Bonham J, Dick DJ, Candy JM, Edwardson JA, Fairbairn A. Cholinergic receptors in cognitive disorders. *Can J Neurol Sci* 1986;13:521-527.
- [172] Rossor MN, Iversen LL, Johnson AJ, Mountjoy CQ, Roth M. Cholinergic deficit in frontal cerebral cortex in Alzheimer's disease is age dependent. *Lancet* 1981;2:1422.
- [173] Francis PT, Palmer AM, Snape M, Wilcock GK. The cholinergic hypothesis of Alzheimer's disease: a review of progress. *J Neurol Neurosurg Psychiatry* 1999;66:137-147.
- [174] George AJ, Holsinger RM, McLean CA, Tan SS, Scott HS, Cardamone T, Cappai R, Masters CL, Li QX. Decreased phosphatidylethanolamine binding protein expression correlates

- with Abeta accumulation in the Tg2576 mouse model of Alzheimer's disease. *Neurobiol Aging* 2006;27:614–623.
- [175] Bader Lange ML, St Clair D, Markesbery WR, Studzinski CM, Murphy MP, Butterfield DA. Age-related loss of phospholipid asymmetry in APP(NLh)/APP(NLh) × PS1-(P264L)/PS-1(P264L) human double mutant knock-in mice: relevance to Alzheimer disease. *Neurobiol Dis* 2010;38:104–115.
- [176] Hamajima N, Matsuda K, Sakata S, Tamaki N, Sasaki M, Nonaka M. A novel gene family defined by human dihydropyrimidinase and three related proteins with differential tissue distribution. *Gene* 1996;180:157–163.
- [177] Kato Y, Hamajima N, Inagaki H, Okamura N, Koji T, Sasaki M, Nonaka M. Post-meiotic expression of the mouse dihydropyrimidinase-related protein 3 (DRP-3) gene during spermiogenesis. *Mol Reprod Dev* 1998;51:105–111.
- [178] Goshima Y, Nakamura F, Strittmatter P, Strittmatter SM. Collapsin-induced growth cone collapse mediated by an intracellular protein related to UNC-33. *Nature* 1995;376:509–514.
- [179] Wang FS, Wolenski JS, Cheney RE, Mooseker MS, Jay DG. Function of myosin-V in filopodial extension of neuronal growth cones. *Science* 1996;273:660–663.
- [180] Arimura N, Inagaki N, Chihara K, Menager C, Nakamura N, Amano M, Iwamatsu A, Goshima Y, Kaibuchi K. Phosphorylation of collapsin response mediator protein-2 by Rho-kinase. Evidence for two separate signaling pathways for growth cone collapse. *J Biol Chem* 2000;275:23973–23980.
- [181] Fukata Y, Itoh TJ, Kimura T, Menager C., Nishimura T, Shiromizu T, Watanabe H, Inagaki N, Iwamatsu A, Hotani H, Kaibuchi K. CRMP-2 binds to tubulin heterodimers to promote microtubule assembly. *Nat Cell Biol* 2002;4:583–591.
- [182] Gu Y, Ihara Y. Evidence that collapsin response mediator protein-2 is involved in the dynamics of microtubules. *J Biol Chem* 2000;275:17917–17920.
- [183] Yoshida H, Watanabe A, Ihara Y. Collapsin response mediator protein-2 is associated with neurofibrillary tangles in Alzheimer's disease. *J Biol Chem* 1998;273:9761–9768.
- [184] Baloyannis SJ, Costa V, Mauroudis I, Psaroulis D, Manolides SL, Manolides LS. Dendritic and spinal pathology in the acoustic cortex in Alzheimer's disease: morphological and morphometric estimation by Golgi technique and electron microscopy. *Acta Otolaryngol* 2007;127:351–354.
- [185] Butterfield DA, Hensley K, Cole P, Subramaniam R, Aksenov M, Aksenova M, Bummer PM, Haley BE, Carney JM. Oxidatively induced structural alteration of glutamine synthetase assessed by analysis of spin label incorporation kinetics: relevance to Alzheimer's disease. *J Neurochem* 1997;68:2451–2457.
- [186] Hensley K, Hall N, Subramaniam R, Cole P, Harris M, Aksenov M, Aksenova M, Gabbita SP, Wu JF, Carney JM, Butterfield DA. Brain regional correspondence between Alzheimer's disease histopathology and biomarkers of protein oxidation. *J Neurochem* 1995;65:2146–2156.
- [187] Carney JM, Carney AM. Role of protein oxidation in aging and in age-associated neurodegenerative diseases. *Life Sci* 1994;55:2097–2103.
- [188] Fukuyama H, Ogawa M, Yamauchi H, Yamaguchi S, Kimura J, Yonekura Y, Konishi J. Altered cerebral energy metabolism in Alzheimer's disease: a PET study. *J Nucl Med* 1994;35:1–6.
- [189] Ogawa M, Fukuyama H, Ouchi Y, Yamauchi H, Kimura J. Altered energy metabolism in Alzheimer's disease. *J Neurol Sci* 1996;139:78–82.
- [190] Gotz ME, Kunig G, Riederer P, Youdim MB. Oxidative stress: free radical production in neural degeneration. *Pharmacol Ther* 1994;63:37–122.

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